

Crowned-coumarin as a New Fluorescence Derivatization Reagent for Carboxylic Acids

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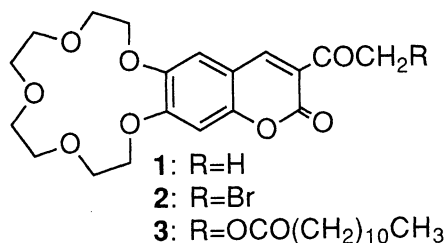
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3-Bromoacetyl-6,7-[2',3'-(1',4',7',10',13'-pentaoxacyclopentadeca)-2'-ene]-coumarin (**2**) was prepared as a new fluorescence derivatization reagent for carboxylic acids in high-performance liquid chromatographic analysis. The derivatization of lauric acid with **2** was readily accomplished under a quite mild condition to yield a highly fluorescent lauric acid derivative. The reaction was accelerated by combining crown moiety as a catalytic site to coumarin fluorophore.

The determination of carboxylic acids in biological samples is significant for the elucidation of various physiological functions in the body. For the purpose, a number of fluorogenic compounds have been developed as fluorescence derivatization reagents for carboxylic acids in high-performance liquid chromatographic (HPLC) analysis.¹⁾ Among these reagents, coumarin derivatives are known to have excellent fluorogenic characters.^{1a-e)} We recently developed 3-bromoacetyl-6,7-methylenedioxy coumarin (BrAMDC) which readily reacts with fatty acids in the presence of 18-crown-6 ether and potassium bicarbonate to give highly fluorescent fatty acid derivatives.²⁾

In the present communication, this derivatization reaction was further extended to simplify and to accelerate the reaction by permitting the reagent itself to hold a certain function. Thus, a crowned-coumarin (**2**) which combined benzo-15-crown-5 (B15Cr5) moiety as

a catalytic site to coumarin fluorophore was designed and prepared as a new fluorescence derivatization reagent for carboxylic acids.



Reagent **2** was prepared from a known compound, 4'-hydroxybenzo-15-crown-5,³⁾ as follows. The Gattermann formylation of starting material in dry ether gave 4'-formyl-5'-hydroxybenzo-15-crown-5 in 73% yield.⁴⁾ This compound was obtained as pasty product, so that it was used for the next step without further purification. Cyclization of the compound was accomplished by refluxing with ethyl acetoacetate and piperidine in ethanol⁵⁾ to give **1** almost quantitatively (94% yield), followed by bromination with tetrabutylammonium tribromide in dichloromethane according to the method of Kajigaeshi *et al.*⁶⁾ gave the desired **2** as yellow needles in 85% yield. The structures of these compounds were confirmed by ¹H-NMR and mass spectral data.⁷⁾

Figure 1 shows the fluorescence spectral responses for **1** in methanol on addition of alkali metal acetates. The fluorescence intensity of **1** was successively decreased in order of $\text{Li}^+ < \text{Cs}^+ < \text{Rb}^+ < \text{K}^+ < \text{Na}^+$ along with the blue shifts of fluorescence maxima, indicating the complexation of the crown site of **1** with metal ions. The stability constants ($\log K_s$) for complexation with K^+ and Na^+ were 2.64 and 2.15, respectively, which were almost comparable with the values of 2.8 and 3.05 for B15Cr5.⁸⁾ From these results, **1** would likely have a catalytic function such as 18-crown-6 ether in BrAMDC derivatization system.

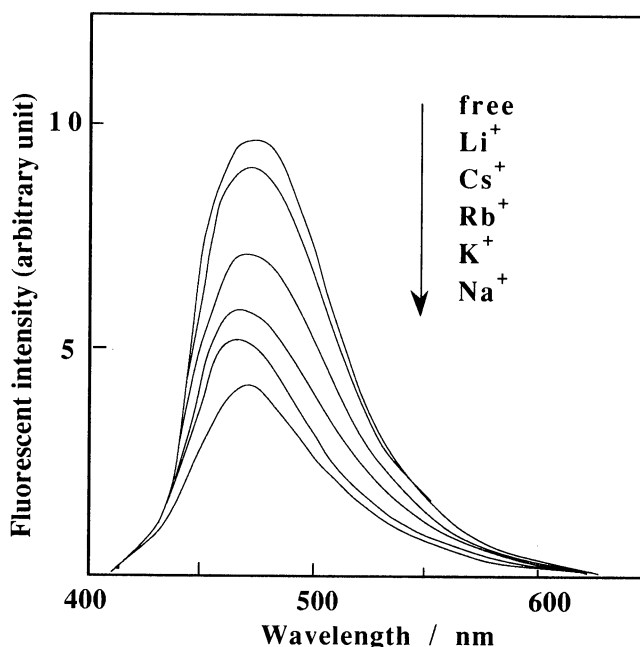


Fig. 1. Fluorescence spectra of **1** in the absence and the presence of alkali metal acetates. $[\mathbf{1}] = 5.0 \times 10^{-6} \text{ mol dm}^{-3}$, $[\text{alkali metal acetate}] = 2.5 \times 10^{-3} \text{ mol dm}^{-3}$

Thus, the derivatization reaction of a test compound, lauric acid, with **2** was attempted in acetonitrile at 30°C in the presence of the bases (KHCO_3 and NaHCO_3) alone and monitored by a reversed phase HPLC equipped with a fluorescence detector ($\lambda_{\text{ex}} = 392 \text{ nm}$, $\lambda_{\text{em}} = 482 \text{ nm}$) using methanol as a mobile phase. The reaction kinetics of **2** with large excess of lauric acid are shown in Fig. 2 together with the results on BrAMDC in the presence of B15Cr5 ether and bases for the comparison. Interestingly, the derivatization of lauric acid were definitely accelerated by the presence of KHCO_3 , but not NaHCO_3 . This finding is matched with the facts that B15Cr5 forms 2:1 complex with K^+ in sandwich manner and 1:1 complex with Na^+ and its complexation ability

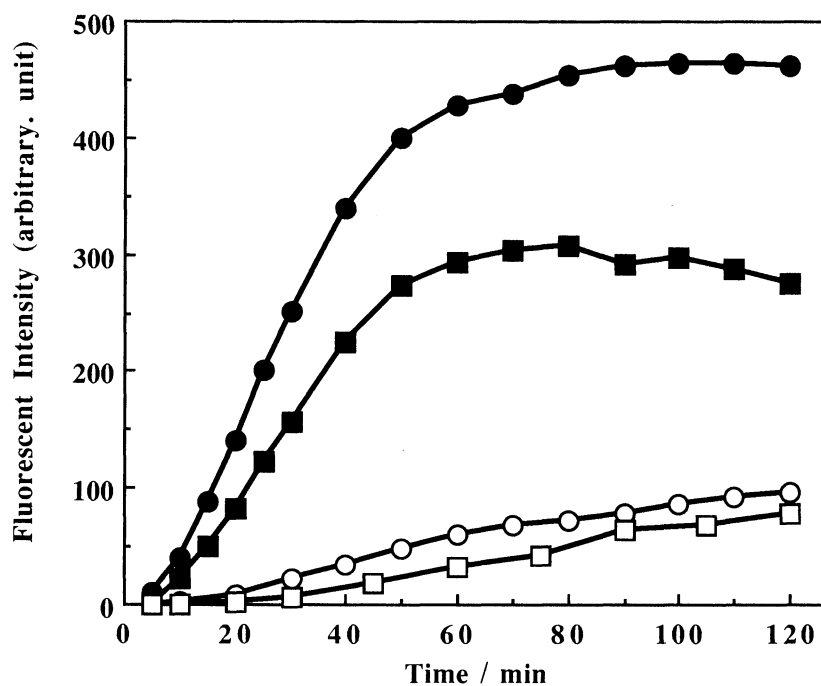


Fig. 2. Time course of the derivatization of lauric acid with **2** and BrAMDC in acetonitrile at 30 °C.

[**2** and BrAMDC] = 2.5×10^{-5} mol dm⁻³, [B15Cr5] = 2.5×10^{-5} mol dm⁻³, [lauric acid] = 2.5×10^{-3} mol dm⁻³, [base] = 2 mg in a reaction solvent (2 ml).

● : **2** + KHCO₃; ■ : BrAMDC + B15Cr₅ + KHCO₃
 ○ : **2** + NaHCO₃; □ : BrAMDC + B15Cr₅ + NaHCO₃

to K⁺ in methanol are slightly greater than that to Na⁺.⁸) The fluorescence quantum yield ($\phi=0.82$) of the derivatized laurate (**3**)⁹ in methanol was very close to the value ($\phi=0.80$) for AMDC-laurate.²) Therefore, the reactivity between **2** and BrAMDC in the presence of KHCO₃ may be approximately comparable by their pseudo first-order rate constants ($k_{\text{obsd}} / \text{Ms}^{-1}$), 8.65×10^{-4} for **2** and 5.31×10^{-4} for BrAMDC, estimated from the lines in Fig. 2. This indicates that **2** apparently surpasses BrAMDC in the reactivity.

From these results, it was suggested that **2** complexes with K⁺ in the same manner as that of B15Cr₅ and that the derivatization reaction with **2** is consequently accelerated because of easy access of laurate anion and reagent molecule.

This is the first example applied a crowned-type reagent to the derivatization reaction for HPLC analysis. The limited data may afford a clue to develop such functionalized reagents for not only carboxylic acids but also other applications. Further works on the other crowned-reagents along this line are currently under way.

References

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- 7) Spectral data and others are as follows. Formyl-compound: $^1\text{H-NMR}$ (CDCl_3) δ 3.73-4.15 (m, 16H, -OCH₂CH₂O-), 6.45 (s, 1H, ArH), 7.05 (s, 1H, ArH), 9.66 (s, 1H, CHO), 11.34 (s, 1H, OH); MS $m/z=312(\text{M}^+)$. **1**: mp 194-196°C; $^1\text{H-NMR}$ (CDCl_3) δ 2.71 (s, 3H, COCH₃), 3.75-4.21 (m, 16H, -OCH₂CH₂O-), 6.80 (s, 1H, C₈-H), 6.97 (s, 1H, C₅-H), 8.46 (s, 1H, C₄-H); MS $m/z=378(\text{M}^+)$. **2**: mp 188-190°C; $^1\text{H-NMR}$ (CDCl_3) δ 3.75-4.23 (m, 16H, -OCH₂CH₂O-), 4.77 (s, 2H, COCH₂Br), 6.81 (s, 1H, C₅-H), 6.99 (s, 1H, C₈-H), 8.58 (s, 1H, C₄-H); MS $m/z=456(\text{M}^+)$, 458 (M^++2).
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- 9) mp 173-5 °C; $^1\text{H-NMR}$ (CDCl_3) δ 0.88 (t, 3H, -CH₃), 1.27-2.49 (m, 20H, -CH₂-), 3.75-4.22 (m, 16H, -OCH₂CH₂O-), 5.37 (s, 2H, C₃-COCH₂O-), 6.81 (s, 1H, C₅-H), 6.98 (s, 1H, C₈-H), 8.56 (s, 1H, C₄-H); MS $m/z=576(\text{M}^+)$.

(Received February 18, 1993)